Indoor aeromycobiota of monumental sites in Minia Governorate

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Abstract. The indoor aeromycobiota contaminating 45 monumental places in Minia Governorate were surveyed during the period from July 2008 to August 2009. The sedimentation plate method along with potato dextrose agar (PDA) medium containing 65 ppm rose bengal were used. A total of 56 fungal species related to 28 genera were obtained from all sampling sites which were chosen to cover four historic eras (16 Pharaonic, 9 Greek Roman, 8 Coptic and 12 Islamic). The major fungal components were *Aspergillus* (10 species), *Penicillium* (5 species), *Alternaria* (3 species), *Cladosporium* (2 species), *Mucor* (3 species), *Ulocladium* (3 species) and *Phoma* (1 species). These fungi are thought to be responsible for the biodeterioration of limestone and woody elements of these historic places.

Key words: Monumental places, aeromycobiota, Minia, Egypt.

Introduction

Studies on indoor aeromycobiota have attracted the attention of several aerobiologists (Arya et al. 2001, Gregory 1973, Sahab et al. 2003, Zielinska-Jankiewicz et al. 2008). The connection between molds and cultural heritage has a long history which includes some myths and mysteries. The so-called 'curse of the pharaoh' - the death of several archaeologists in the team of Howard Carter after discovery and opening of Tutankhamun's tomb - was later explained by the fact that spores of the pathogenic fungi Aspergillus niger and Aspergillus flavus were found on and in the sarcophagi and that a lung infection or other systemic mycosis - aspergillosis - was the possible reason for death. Molds always were and still are threatening historical and contemporary material of objects of art in libraries and in museums (Pangallo et al. 2009, Sterflinger 2010). Fungi are also found on mural paintings in churches, in caves and in catacombs and even as biodeteriogens of architectural surfaces and stone monuments (Ettenauer et al. 2010, Steiger et al. 2011). The oldest and most precious objects suffering from serious fungal invasions are rock art caves as for example Lascaux in France (Bastian and Alabouvette 2009). Fungi cause serious esthetical spoiling of paintings, sculptures, costumes, ceramics, mummies, books and manuscripts. Due to their ability to form hyphal networks they penetrate materials deeply, resulting in material loss due to acid corrosion, enzymatic degradation and mechanical attack. Fungi in libraries, museums and their storage rooms can seriously threaten the health of the restorers, of the museum personnel and of the visitors due to their allergic

potential, the production of mycotoxins and their ability to cause systemic infections in humans (Crook and Burton 2010). Airborne fungal spores in storage rooms of museums can well reach levels of more than 8000 per m³. According to Yang and Johanning (1997) and Urzi and Realini (1998), transport and ultimate settling on the surfaces is affected by: i) the physical properties of particles and/or of droplets (size, density and shape); ii) the environmental parameters (magnitude of air movement, relative humidity and temperature) and iii) the bioreceptivity of the surface itself and nutrient availability. In Egypt, El-Hissy et al. (1991) studied the mycobiota of water and mud samples from sacred lake and wells at Karnak Temple (Luxor), Dandara Temple (Qena) and Abidos Temple (Sohag)). The common fungi in these historic places were Alternaria alternata, Aspergillus flavus, A. fumigatus, A. niger, A. terreus. Other fungal species belonging to Cladosporium, Fusarium, Mucor, Penicillium, Rhizopus, Ulocladium were also recorded. However, no available records on the airborne mycobiota of monumental places in Egypt and their impact on monuments, rocks, wooden blocks, mummies, as well as on health status of workers and visitors. This study was undertaken to to isolate and identify the indoor airborne fungi in monumental sites related to four historic eras (Pharaonic, Greek Roman, Coptic and Islamic) distributed in Minia Governorate.

Materials and Methods

The airborne fungal spores were surveyed in 45 monumental places belonging to 4 historic eras distributed in Minia Governorate as follows (16 Pharaonic, 9 Greek Roman, 8 Coptic and 12 Islamic eras) (Fig. 1, Table 1). The study was extended for 14 months from July 2008 to August 2009. The exposed plate method (Hoekstra et al. 2004) was employed to trap the fungal propagules contaminating the air inside these places. Three plates (9 cm diam.) containing sterile potato dextrose agar (PDA) with rose bengal (65 ppm) as a bacteriostatic agent were used. The sampling time was set for 30 minutes at an approximate height of 50 cm. from the floor. After trapping, cultures were incubated at 28°C for 7-10 days. The growing fungi were counted, isolated and identified. Non- sporulating and slow-growing fungi were transferred to other media such as Czapek's Dox agar, PDA and potato sucrose agar (PSA) without rose bengal to allow better growth and sporulation. The counts were expressed as colony-forming units (CFUs) / 3 plates in individual exposure.

Identification of the trapped fungi

Identification of fungal genera and species was done on the basis of morphological characteristics using taxonomic keys and descriptions given by Raper and Fennell (1965), Pitt (1979), Sivanesan (1987), Moubasher (1993, Leslie and Summerell (2006) and Domsch *et al.* (2007).



Fig.1: Map of monumental sites visited in Minia Governorate

Sample	Site and place	Era	District
N0.			
1	El-Qayatty mosque	Islamic	El-Edwa
2	El-Snouldamy mosque (Aba El-waqi)	Islamic	Magnagna
3	Marry Gerges church (Ashnien)	Coptic	Maghagha
4	El Harry El Caldanarge (El Daharry)	Latentia	
5	El-Hassan El-Salen mosque (El-Bannassa)	Islamic	Bani- Mazar
0	SedyAly El-Galilani alea (El-Balmassa)	Islamic	Bani- Mazar
8	Abu- Samra mosque (El-Bahnassa)	Islamic	Bani- Mazar
9	Abu-Samraminarte (El-Bahnassa)	Islamic	Bani-Mazar
10	El-SabaaBannat region (El-Bahnassa)	Islamic	Bani-Mazar
11	Virgin Mary church (Bardanoha village)-Orthozex church	Coptic	Mattay
12	Ava Oastour church (Bardanoha village)-Orthozex church	Coptic	Mattav
13	Virgin Mary church (Gabal El-Teir)	Coptic	Samalott
14	The two monumental doors area –God Hathour closet- Merembtah area	Pharaonic	Samalott
	(El-Saryrya)		
15	El-Lamatty mosque (Minia city)	Islamic	Minia
16	El-Amrawy mosque (Minia city)	Islamic	Minia
17	Nero Temple (Tehna El-Gabal)	Greek- Roman	Minia
18	Embalming area (Tehna El-Gabal)	Greek- Roman	Minia
19	The God Hathour room (Tehna El-Gabal)	Greek-Roman	Minia
20	Crocodile room (Tehna El-Gabal)	Greek-Roman	Minia
21	Anka-ankh tomb (Frezer tombs) (El-Hawarta village)	Pharaonic	Minia
22	Kahab tomb (Frezer tombs) (El-Hawarta village)	Pharaonic	Minia
23	Hnoka tomb (Frezer tombs) (El-Hawarta)	Pharaonic	Minia
24	Saint Abahour church (Sawada village)	Coptic	Minia
25	The Pyramide base (Zaweit Sultan village)	Pharaonic	Minia
26	KhenmHotop II tomb (Bani-Hassan El-Shorouq village)	Pharaonic	Abu-Qurqas
27	Khety tomb (No.17) (Bani-Hassan El-Shorouq village)	Pharaonic	Abu-Qurqas
28	Bakht tomb (Bani-Hassan El-Shorouq village)	Pharaonic	Abu-Qurqas
29	Amenemhattomb (No.2) (Bani-HassanEl-Shorouq village)	Pharaonic	Abu-Qurqas
30	Pakhttemple Queen Hatshepsut and Tuhtmos III temple (IstableAntar)	Pharaonic	Abu-Qurqas
31	Embalming room (IstableAnter)	Pharaonic	Abu Qurqas
32	El-Yousfy mosque (Mallawi city)	Islamic	Mallawi
33	El-Asqalany mosque (Mallawi city)	Islamic	Mallawi
34	Bazilica church (El-Ashmonein)	Greek-Roman	Mallawi
35	Petosiris tomb (Tuna El-Gabal)	Greek-Roman	Mallawi
36	Isadora tomb (Tuna El-Gabal)	Greek- Roman	Mallawi
37	Catacombs of Ibis and Baboons (Tuna El-Gabal)	Greek-Roman	Mallawi
38	Ramsis II Temple (El-Sheikh Ebada)	Greek-Roman	Mallawi
39	Anba Beshoy church (Deir El-Barsha)	Coptic	Mallawi
40	Abu-Hennis church (Deir Abu-Hennis)	Coptic	Mallawi
41	Pentu tomb (No.5) (Tell El-Amarna) (Ekhat-Aton)	Pharaonic	DierMawas
42	MerireI tomb (No.4) (Tell El-Amarna)	Pharaonic	DierMawas
43	Ahmos tomb (No.3) (Tell El-Amarna)	Pharaonic	DierMawas
44	Aye tomb (No.25) (El-Haj Qandel)	Pharaonic	DierMawas
45	Aye tomb (No.23)(El-Haj Qandel)	Pharaonic	DierMawas

Table 1: Sites from which airborne fungi were sampled.

Results and Discussion

Fifty- seven species belonging to twentyeight genera were collected from the indoor air samples in all different places of the four historical places as shown in Table (2). *Aspergillus* was consistently the most frequent genus (100% of exposures) and contributed the broadest spectrum of species (10 species) of which *A. niger* and *A. flavus* were of high frequency (33 and 23 sites out of 45) contributing 38.42% and 8.61% of the total count respectively.

Pencillium was moderately recovered from the indoor air of 14 monumental places matching 8.8% of total fungal catch. It was represented by six species of which *P. chrysogenum* and *P. purpurogenum* were common sharing with 3.14% and 4.19% of the total count respectively.

Alternaria and Cladosporium appeared in moderate incidence in the air of monumental places (17 sampling sites for each) matching 4.6 % and 17.0% of total fungal catch. Alternaria was represented by species of which A. alternata was the basic constituent (14 sampling sites out of 45). Cladosporium was represented by two species, namely C. herbarum and C. sphaerospermum which appeared in 9 and 8 places contributing for 12.96% and 4.61% of total fungal catch respectively. The rest of genera and species were isolated in low frequency with counts ranging from 1.26% to 0.21% of the total count.

Comparing the indoor aeromycobiota of the individual historic eras, results in Table (3) revealed that the highest fungal counts (260 CFU) and the broadest spectra of fungal genera (21) and species (40) were obtained from the monumental sites belonging to the Pharaonic era. This might be due to the relatively greater number of sampling sites of that era (16 sites) compared to limited number of sites belonging to other historic eras. The richest places in aeromycobiota were the Pharaonic tombs Pentu (No. 41) and Khnem-Hotop-II (No.26) located respectively at Tell El-Amarna and Bani Hassan El-Shoroug villages (which yielded nine fungal species for each). The poorest place was the Pharaonic tomb of Bakhet (Abu-Ourgas from which A. flavus was the only isolated fungus). Aspergillus was the most dominant genus in the air of Pharaonic places. It was represented by eight species and contributed 67.3% of the total count. A. niger and A. flavus constituting 34.40% and 25.60% of the total count respectively. The other species were identified in low frequency. Alternaria occurred also in high incidence contaminating 50% of sampling sites accounting for 4.6% of total fungi. Among the three species of Alternaria, A. alternata was the major representative (50% of samples matching 4.6% of total fungi). Penicillium, Cladosporium, Mucor and Ulocladium occurred (25%-43.8% in moderate of samples contributing 2.0 %-8.8% of total fungal catch). Penicillium was represented by four species of which P. chrysogenum and P. purpurogenum were moderately encountered (25% of samples for each) matching 1.6% and 5.8% of total fungi respectively. The remaining genera and species were less frequently isolated from the indoor air of the monumental places belonging to the Pharaonic era.

Regarding the Greek Roman era, results in Table (3) indicated that the lowest fungal population (68 CFU) and the narrowest spectrum of fungal genera (7) and species (9) were isolated from the 9 monumental sites of this era. The richest sampling site was Isadora tomb which yielded 6 species whereas the poorest place was Catacombs which yielded only Penicillium italicum. The most common genus was Aspergillus which was recovered from all places contributing 73.5% of the total count. It was represented by A. niger and A. flavus which appeared in 4 and 5 sites out of 9 matching 16.7% and 47.6% of total fungal population respectively. Each of Alternaria and Cladosporium appeared in 33.3% of sampling sites accounting respectively for 13.2% and 8.8% of total fungal catch. A. alternata and A. tenuissima were only represented in 11.1% and 22.2% of air samples matching 4.2% and 9.0% of total fungi respectively. C. herbarum and C. sphaeropermum occurred in 33.3% and 11.1% of samples representing 5.1% and 2.6% of total fungi respectively.

From the monumental places of Coptic era, 83 fungal colonies were obtained and classified in 19 species belonging to 13 genera. Aspergillus was the most common genus (100% of samples) and was represented by four species comprising 32.5% of the total count. A. flavus was represented in 50% of air samples sharing in the total count with 7.2%. Each of Cladosporium and Phoma appeared in moderate incidence (37.5% of indoor air samples) matching respectively 37.5% and 3.5% of total fungal catch. Penicillium, Alternaria and Mucor occurred also in moderate incidence (25% of sampling sites for each) whereas the remaining genera and species were recovered in low frequency and in low counts. The highest number of fungal species was identified in Saint Abahour church (11 species) whereas the lowest number (one species) was isolated from Dier Abu Henes church which was identified as A. niger.

Table 2: Indoor aeromy	ycobiota recovered from the 4	5 monumental places in Minia	a Governorate on PDA medium
at 28°C.			

Genera & species	Total count	Total count %	Frequency	O. R.
Aspergillus	252	52.83	45	Н
A. candidus Link	2	0.41	2	L
A. clavatus Desmazieres	1	0.20	1	L
A. flavipes (Bain. & Sart.) Thom & Church	4	0.83	1	L
A. flavus Link	41	8.59	23	Н
A. niger van Tieghem	182	38.15	33	Н
A. fumigatus Fresenius	6	1.25	6	L
A. ochraceus Wilhelm	8	1.67	1	L
A. terreus Thom	4	0.83	4	L
A. ustus (Bain.) Thom & Church	4	0.83	2	L
Penicillium	42	8.80	14	М
P. chrysogenum Thom	15	3.14	5	L
P. funiculosum Thom	2	0.41	2	L
<i>P. italicum</i> Wehmer	5	1.04	6	L
P nurpurogenum Stoll	20	<u>/ 10</u>	7	T
	20	4.17	17	
Alternaria	22	4.61	1/	M
A. alternata (Fr.) keissler	19	3.98	14	M
A. chlamydospora Mouchacca	1	0.20	1	L
A. tenuissima (Kurze ex. Pers.)Wiltshire	2	0.41	2	L
Acremonium rutilum W.Gams	1	0.20	1	L
Botryodiplodia theobromae Patouillard	1	0.20	1	L
Cladosporium	81	17.00	17	М
C. herbarum (Pers.) Link: Fr.	59	12.36	9	L
C. sphaerospermum Penzig	22	4.61	8	L
Cochliobolus	5	1.03	6	L
C. bicolor Paul & Parbery	1	0.20	1	L
C. lunatus (Shear) Boedijn	4	0.83	5	L
Chaetomium	3	0.62	2	L
C. fimicola Cooke	2	0.41	1	L
C. indicum Corda	1	0.20	1	L
Drechslera erythrospila (Drechsler) Shoemaker	1	0.20	1	L
Doratomyces stemonitis (Persoon) Morton & Smith	1	0.20	1	L
Emericella nidulans var. lata (Eidam) Vuill.	1	0.20	1	L
Phoma epicoccina Punithalingam. Tulloch & Leach	9	1.88	7	L
Eurotium amstelodami Mangin	1	0.21	1	L
Fusarium	4	0.83	2	L
<i>F</i> chlamydosporum Wollenw & Reninking	2	0.41	1	L
<i>F. semitectum</i> Berk. & Ray.	2	0.41	1	L
Humicola grisea Traaen	1	0.20	1	L
Mucor	11	2.30	8	L
<i>M. circinelloides</i> van Tiegh.	2	0.41	1	L
<i>M. fuscus</i> Bainier	1	0.20	1	L
<i>M. racemosus</i> Fresenius	8	1.67	5	L
Malbranchea sulfurea (Miehe) Coony & Emerson	1	0.20	1	L
Nigrospora sphaerica (Sacc.) Mason	2	0.41	2	L
Oidiodendron chlamvdosporum Robak	2	0.41	2	L
Papulaspora irregularis Hoston	1	0.20	1	L
Paecilomyees variotii Bainier	1	0.20	1	L
Rhizopus oryzae Went & Geerl.	3	0.63	2	L
Setosphaeria. rostrata (Drechsler) Subram. & Jain	3	0.63	3	L
Stachybotrys chartarum (Ehrenberg) Hughes	8	1.67	9	L

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Genera & species	Total count	Total count %	Frequency	O. R.
Stemphylium	2	0.41	2	L
S. botryosum Wallroth	1	0.20	1	L
S. vesicarium (Wallroth) E. G. Simmons	1	0.20	1	L
Syncephalastrum racemosum Cohn ex Schröter	1	0.20	1	L
Trichoderma harzianum Rifai	9	1.89	4	L
Tritirachium oryzae (Vincens) de Hoog	1	0.20	1	L
Ulocladium	7	1.46	5	L
U. botrytis Preuss	3	0.62	3	L
U. chlamydosporum Mouchacca	3	0.62	3	L
U. chartarum (Preuss) E. G. Simmons	1	0.20	1	L
Gross total count (CFU)	477			
Total No. of genera	28			
Total No. of species	52			

Occurrence remarks: High (H) = 23 - 45 sites, Moderate (M) = 11 - 22 sites and Low (L) = Less than 11 sites.

When the aeromycobiota of the 12 monumental places of Islamic era were surveyed it was observed that these sites were contaminated with twenty-seven species belonging to sixteen genera with a total fungal catch of 136 CFU. Aspergillus was also the most common genus contaminating the air of all sites. It was represented by six species contributing 63.0% of the total count. A. niger and A. flavus were also the most prevalent species being recovered from 100% and 50% of sites comprising 42.22% and 16.3% of the total fungal count respectively. Penicillium, Alternaria, Cladosporium and Stachybotrys occurred in moderate incidence (25%-41.7% of samples accounting for 3.7%-10.4% of total fungal population). The richest sample was recorded in El-Qayatty mosque which contained eight species. Two places were considered to be the poorest which were El-Yousfey mosque (Mallawi City) and El-Seidah Rouqaya area (Bani Mazar City) from which Trichoderma harzianum and A. niger were respectively isolated.

Results of the present investigation showed great similarity with those obtained from different monumental sites around the world. In Italy, Urzi et al. (2001) studied the air borne fungi of Messina museum and found that Aspergillus, Penicillium, Fusarium, Alternaria, Cladosporium, Ulocladium, Aureobasidium and *Phoma* were the most common. The authors mentioned that once fungi settle on and colonize stone surfaces they contribute to a great variety of alterations like black patinas, inter-granular growth, marble sugaring, biopitting, etc. In Roman catacombs, Saareia et al. (2004) isolated fungi from both air and biofilm samples namely Lecanicillium psalliotae, L. aranearum, Torrubiella spp. and Beauveria alba and less frequent fungi include Alternaria spp., Cladosporium spp. and Curvularia spp. In

China, Wanfu (2010) studied fungal spores in open and closed caves from September 2008 to August 2009, the most prevalent species were related to Cladosporium, Penicillium, Alternaria and Aspergillus. In Turkey, Nuhoglu (2006) studied the accelerating effects of microorganisms on the biodeterioration of stone on monuments under air pollution aged from 441 to 823 years old. The results showed that the microorganisms could develop during the winter months when the night time average temperature was even -25°C.

In non-historic places the indoor mycobiota showed also high similarity with those identified in the present investigation. In Madrid (Spain), Silvia et al. (2007) made an aero mycological sampling during 2003 and 2004 in atmosphere of the University Campus. They were able to identify 70 fungal species in 2003 and 83 species in 2004, the most abundant were C. cladosporioides, C. herbarum, and unknown species of Aspergillus, Pleospora and Bovista. In Romania, Nicoleta and Dorina (2009) recorded the most prevalent fungal spores in the air were species of Cladosporium, Setosphaeria, Alternaria and Epicoccum. In Italy, Anna and Marinella (2000) surveyed the air spora of two Milan underground stations and found that Cladosporium, Penicillium, Epicoccum and Alternaria were the common genera.

The expected source of indoor contamination by fungi is the movement of outdoor fungal propagules from open fields, vegetation, soil, deteriorating plants and others to thes monumental places. This expectation can be supported by the great similarity between these fungal species and those previously isolated by Mazen and Shaban (1983) from wheat fields in Minia Governorate. During that study Aspergillus spp. (A. terreus, A. niger and A. flavus), Alternaria alternata, Cladosporium spp. (C. cladosporiodes and C. herbarum) and

55

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Eras	Pharaonic era (16 sites)			Greek Roman era (9 sites)					Coptic era (8 sites)					Islamic era (12 sites)						
Genera & species	TC	%TC	F	%F	OR	TC	%TC	F	%F	OR	TC	%TC	F	%F	OR	TC	%TC	F	%F	OR
Aspergillus	175	67.3	16	100	Η	50	73.5	9	100	Н	27	32.5	8	100	Н	85	63.0	12	100	Η
A. candidus	1	0.4	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	0.7	1	8.3	L
A. clavatus	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	1.2	1	12.5	L	0	0.0	0	0.0	0
A. flavipes	1	0.4	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
A. flavus	64	25.6	9	56.3	Μ	13	16.7	4	44.4	Μ	6	7.2	4	50.0	Μ	22	16.3	6	50.0	Η
A. fumigatus	4	1.6	3	18.8	L	0	0.0	0	0.0	0	1	1.2	1	12.5	L	2	1.5	2	16.7	L
A. niger	86	34.4	11	68.8	Η	37	47.4	5	55.6	Н	19	22.9	5	62.5	Н	57	42.2	12	100	Η
A. ochraceus	2	0.8	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
A. terreus	3	1.2	3	18.8	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	2	1.5	1	8.3	L
A. ustus	14	5.6	1	6.6	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	0.7	1	8.3	L
Penicillium	23	8.8	7	43.8	Μ	10	14.7	1	11.1	L	8	9.6	2	25.0	М	14	10.4	4	33.3	М
P. chrysogenum	4	1.6	4	25.0	Μ	0	0.0	0	0.0	0	0	0.0	0	0.0	0	2	1.5	1	8.3	L
P. funiculosum	1	0.4	2	6.6	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
P. italicum	3	1.2	2	12.5	L	10	12.8	1	11.1	L	2	2.4	1	12.5	L	3	1.20	2	12.5	L
P. purpurogenum	15	68.	4	25.0	М	0	0.0	0	0.0	0	6	7.1	1	12.5	L	9	6.7	2	12.5	L
Alternaria	12	64.	8	50.0	Η	9	13.2	3	33.3	М	4	4.8	2	25.0	М	5	3.7	4	33.3	М
A. alternata	10	4.0	8	50.0	Η	2	4.2	1	11.1	L	4	4.7	2	25.0	М	5	3.7	4	33.3	М
A. tenuissima	1	0.4	1	6.3	L	7	9.0	2	22.2	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0
A. chlamydosporum	1	0.4	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
Acremonium rutilum	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	0.7	1	8.3	1
Botryodipolodia theobromae	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	0.7	1	8.3	L
Chaetomium	1	0.4	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	0.7	1	8.3	L
C .indicum	1	0.4	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
C. fimicola	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	0.7	1	8.3	L
Cladosporium	16	6.2	5	31.3	Μ	6	8.8	3	33.3	М	31	37.3	3	37.5	М	12	8.9	5	41.7	М
C. herbarum	11	4.4	2	18.8	L	4	5.1	3	33.3	М	7	8.3	2	25.0	М	2	1.5	2	12.5	L
C. sphaerospermum	5	2.0	2	12.5	L	2	2.6	1	11.1	L	25	29.4	2	25.0	М	10	7.4	3	25.0	Μ
Cochliobolus	3	1.2	3	18.8	L	3	4.4	2	22.2	L	1	1.2	1	12.5	L	0	0.0	0	0.0	0
C. bicolor	1	0.4	1	6.3	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
C. lunatus	2	0.8	2	12.5	L	3	4.4	2	22.2	L	1	1.2	1	12.5	L	0	0.0	0	0.0	0

Table 3: Indoor aeromycobiota recovered on PDA at 28° C from the different monumental places

Table 3: Continued Coptic era (8 sites) Eras Pharaonic era (16 sites) Greek Roman era (9 sites) Islamic era (12 sites) %TC %F TC %TC %F TC %TC %F TC %TC Genera & species TC F OR F OR F OR F %F OR 0 0 Doratomyces stemonitis 0.4 6.3 0.0 0.0 0 0.0 0 0.0 0 0 0.0 0 0.0 0 1 1 L 0 Drechslera.erythrospila 0.4 6.3 0 0.0 0 0.0 0 0 0.0 0 0.0 0 0 0.0 0 0.0 0 1 1 L Emericilla.nidulans var. lata 1 0.4 1 6.3 L 0 0.0 0 0.0 0 0 0.0 0 0.0 0 0 0.0 0 0.0 0 Eurotium.amstelodami 0 0.0 0 0.0 0 0 0.0 0 0.0 0 0 0.0 0 0.0 0 1 0.7 1 8.3 L 1.2 12.5 0.0 Fusarium 1 0.4 1 6.3 L 0 0.0 0 0.0 0 1 1 L 0 0.0 0 0 F. chlamydosporum 0 0 0 0.0 0 0.4 6.3 L 0.0 0.0 0 0 0.0 0.0 0 0 0.0 0 1 1 1.2 12.5 F. semitectum 0.0 0 0.0 0 0 0.0 0 0.0 1 L 0 0.0 0 0.0 0 0 0 1 0.0 8.3 Humicola grisea 0 0.0 0 0.0 0 0 0.0 0 0.0 0 0 0 0.0 0 1 0.7 1 L Malbranchea sulfurea 0 0 0 0.0 0.7 8.3 0 0.0 0 0.0 0 0.0 0.0 0 0.0 0 0 1 1 L 25.0 0.0 2 2.4 25.0 2 1.5 2 12.5 5 4 0 0 0 2 L L Mucor 2.0 L 0.0 M. circinelloides 0 0 0 0 0.0 0 0.0 0 0.7 12.5 L 0 0.0 0 0.0 0.0 0.0 0 1 1 M. fuscus L 0 0 2 2.4 1 12.5 L 0 0.0 0.0 0 1 0.4 1 6.3 0.0 0.0 0 0 4 3 L 0 0.0 0 0 1 1 12.5 L 1 0.7 8.3 L M. racemosus 1.6 18.8 0.0 1.2 1 Nigrospora sphaerica 2 0.8 2 12.5 L 0 0.0 0 0.0 0 0 0.0 0 0.0 0 0 0.0 0 0.0 0 Oiodiodendron 1 0.4 6.3 L 0 0.0 0 0.0 0 0 0.0 0 0.0 0 1 0.7 8.3 L 1 1 chlamydosporum Paecilomyces variotii 6.3 00. 0.0 0.0 0.0 0.0 0.0 1 0.4 L 0 0 0 0 0 0 0 0 0 1 Phoma epicoccina 2 4 5.1 3 3.5 3 37.5 Μ 2 1.5 8.3 2 0.8 12.5 L 1 11.1 L 1 L 1.2 Rhizopus oryzae 1 0.4 6.3 L 0 0.0 0 0.0 0 1 1 12.5 L 0 0.0 0 0.0 0 1 0.0 1.2 12.5 2 1.5 12.5 L Setosphaeria. rostrata 1 0.6 1 6.3 L 0 0.0 0 0 1 1 L 1 1.2 12.5 3.7 2 0 0 5 3 25.0 Μ Stachybotrys chartarum 2 0.8 12.5 L 0.0 0.0 0 1 1 L 0.0 2 2 0 0 0 0 1 0 0 0 Stemphylium 0.8 12.5 L 0.0 0.0 0.0 0.0 0 0.0 S. botryosum 0 0 0 L 1 0.4 1 6.3 L 0.0 0.0 0 0 0.0 0.0 0 1 0.40 1 6.3 S. vesicarium 0 0 0 0 0 0 1 0.4 1 6.3 L 0.0 0.0 0.0 0 0.0 0 0.0 0 0.0 Syncephalastrum 0.0 6.3 0 0 0.0 0 1 0.4 L 0 0.0 0 0 0.0 0 0.0 0 0.0 0 1 racemosum 12.5 8.8 11.1 0.0 0.0 2 L L 0 0 0 Trichoderma harzianum 2 0.8 6 1 1 0.7 1 8.3 L

Table 3: Continued:																				
Eras	Pharaonic era (16 sites)					Greek Roman era (9 sites)					Coptic era (8 sites)					Islamic era (12 sites)				
Genera & species	TC	%TC	F	%F	OR	TC	%TC	F	%F	OR	TC	%TC	F	%F	OR	TC	%TC	F	%F	OR
Tritirachium oryzae	0	0.0	0	0.0	0	0	0.0	0	0.0	0	1	1.2	1	12.5	L	0	0.0	0	0.0	0
Ulocladium	7	2.8	5	31.3	М	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
U. botrytis	2	0.8	3	12.5	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
U. chartarum	3	1.2	3	18.8	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
U. chlamydosporum	2	0.8	2	12.5	L	0	0.0	0	0.0	0	0	0.0	0	0.0	0	0	0.0	0	0.0	0
Total Count (CFUs)	260						68						136							
No. of genera	21					7				13					16					
No. of species	40					9				19					26					

% F= Percentage frequency (calculated per 16 places from Pharaonic era, 9 from Greek Roman era, 8 from Coptic era and 12 from Islamic era); OR= Occurrence remarks; H= 50.0% - 100%, M=25.0% - 49.0%, L= Less than 49.0% (Pharaonic), Counts are calculated as colony- forming units (CFU) / 3 plates and 30 min. exposure each.

Fusarium solani were isolated. In Assuit, Moubasher and Moustafa (1974) isolated *Cladosporium, Alternaria, Aspergillus* and *Penicillium* as common air borne fungi. Also Ismail *et al.* (2010) recorded *Alternaria, Aspergillus, Cladosporium, Eurotium, Penicillium* and *Rhizopus* from the indoor air of the external dermatology clinics of Assuit University Hospitals.

As mentioned by Helmi et al. (2011) biodeterioration of mural paintings by A, niger and A. flavus has been proved in different mural paintings in Egypt. These fungi were observed on mural paintings of Nfer Bau Ptah Tomb, Giza, Egypt. Successful cleaning of these paintings was done with the antibiotic "6penthyl α -pyrone phenol" extacted from Trichoderma sp. followed by fumigation of the tomb by isopropy (alcohols to avoid any new contamination by fungal spores). During their mycological analysis of some Egyptian mummies, Elnagger et al. (2010) were able to isolate Alternaria tenuis, Aspergillus niger, Chaetomium globosum and Penicillium corylophilum. Lopez-Martinez et al. (2007) isolated 469 fungal colonies from 12 mummies deteriorated with fungi. Among the isolated fungi, Penicillium, Cladosporium and Aspergillus were identified. Abdulla et al. (2008) isolated some species of actinomycetes from decayed and sound stone taken from a tomb at Tell Basta, Zagazig City, Egypt.

In conclusion, The results of this study indicated that the air mycoflora of monumental places were almost similar to those obtained from other places. The role of these fungi on deterioration of monumental elements will be studied in the future.

References

- Anna MP and Marinella R (2000): Air-borne fungi as biocontaminants at two Milan underground stations. International Biodeterioration & Biodegradation 45(1-2): 43-47.
- Abdulla H, May E, Bahgat M and Dewedar A (2008): Characterisation of actinomycetes isolated from ancient stone and their potential for deterioration. Polish Journal of Microbiology 57(3): 213-220.
- Arya A, Shab AR and Sadasivan S (2011): Indoor aeromycoflora of Baroda museum and deterioration of Egyptian mummy. Current Science 81(7):793-799.
- Bastian F and Alabouvette C (2009): Lights and shadows on the conservation of a rock art cave: The case of Lascaux cave. International Journal of Speleology 38: 55-60.

- Crook B and Burton NC (2010): Indoor moulds, sick building syndrome and building related illness. Fungal Biology Reviews 24: 1-8.
- Domsch KH, Gams W and Anderson TH (2007): Compendium of soil fungi. 2 ed, IHW-Verlag, Eching. Germany, pp.672.
- El-Hissy FT, Khallil AM and El-Naghy MA (1991): Mycoflora of water pools in the vicinity of some ancient Pharaonic Temples in Upper Egypt. Journal of Islamic Academy of Sciences 4(4): 293-296.
- Elnaggar A, Sahab A, Ismail S, Mahgoub G and Abdelhady A (2010): Micorobial study of Egyptian Mummies: An assessment of enzyme activity, fungicides and some mummification materials for the inhibition of microbial deterioration. E-Conservation Magazine (16): 39-49.
- Ettenauer J, Sterflinger K and Piner G (2010): Cultivation and molecular monitoring of halophilic microorganisms inhabiting an extreme environment presented by a saltattacked monument. International Journal of Astrobiology 9: 59-72.
- Gregory PH (1973): Microbiology of atmosphere, 2nd Ed. Leonard Hill Books, Aylebury, England, pp.377.
- Helmi FM, Elmitwalli HR, Rizk MA and Hagrassy AF (2011): Antibiotic extraction as a recent biocontrol method for *Aspergillus niger* and *Aspergillus flavus* fungi in ancient Egyptian mural paintings. Mediterranean Archaeology and Archaeometry 11(2): 1-7.
- Hoekstra ES, Samson RA and Summerbell RC (2004): Methods for the deterioration and isolation of fungi in the indoor environments. PP. 298-305. In: Samson RA, Hoekstra ES and Frisvad JC, Introduction to food- and air-borne fungi Centraalbureau voor Schimmelcultures, Utrecht, the Netherlands, 7th edition.
- Ismail MA, Moubasher AH, Al-Ryani M and Farhan MA (2010): Seasonal fluctuations of potentially pathogenic indoor aeromycobiota in the external dermatology clinics of Assuit University Hospitals. Journal of Basic and Applied Mycology 1: 11-21.
- Leslie J and Summerll BA (2006): *Fusarium* Laboratory Manual. Blackwell publishing professional; 1st edition, Iowa USA, pp. 388.
- Lopez-Martinez R, Hernandez-Hernandez F, Melan-Chiu BE, Manzano-Gayosso P and Mendez-Tovar LJ (2007): Effectiveness of Imazalil to control the effect of fungal deterioration of mummies at the Mexico

City Museum "El-Carmen" Revesta Iberoamericana de Mycologie 24: 283-288.

- Mazen MB and Shaban GM (1983): Airborne fungi of wheat fields in Egypt. Qatar University Science Bulletin 3:131-139.
- Moubasher AH (1993): Soil fungi in Qatar and other Arab Countries. The Scientific and Applied Research Center, University of Qatar, Doha, pp. 566.
- Moubasher AH and Moustafa AF (1974): Airborne fungi at Assiut, Egypt. Egyptian Journal of Botany 17(2, 3): 135-179.
- Nicoleta I and Dorina T (2009): Aeromycoflora in outdoor environment of Timisoara City. Notulae Scientia Biologicae 1(1): 21-28.
- Nuhoglu Y, Oguz E, Uslu H, Ozbek A, Ipekoglu B, Ocka I and Hasenekoglu I (2006): The accelerating effects of the microorganisms on biodeterioration of stone monuments under air pollution and continental-cold climatic conditions in Erzurum, Turkey. Science of the Total Environment 364 (1-3): 272-283.
- Pangallo D, Chovanova K, Simonovicova A and Ferianc P (2009): Investigation of microbial community isolated from indoor artworks and their environment: identification, biodegradative abilities, and DNA typing. Canadian Journal of Microbiology 55: 277-287.
- Pitt JI (1979): The genus *Penicillium* and its teleomorphic states *Eupenicillium* and *Talaromyces*. Academic Press, London, pp. 634.
- Raper KB and Fennell PI (1965): The genus Aspergillus. Williams and Willims, Baltimore. USA, pp. 686.
- Saariea M, Alakomi HL, Suihko ML, Maunuksela L, Rasska L and Mattila-Sandholm T (2004): Heterotrophic microorganisms in air and biofilm samples from Roman catacombs, with special emphasis on actinobacteria and fungi. International Biodeterioration and Biodegradation 54(1): 27-37.
- Sahab AF, Tawfic F, Sahaba S and Moustafa S (2003): Indoor fungal airospora and microorganisms communities associated with old manuscripts of GEBO of Egypt. Journal of Agricultural Sciences, Mansoura University 28(8): 6055 – 6063.

- Silvia, S, Alberto D and Montserrat G (2007): Monitoring of airborne fungi in Madrid (Spain). Acta Botanica Croatica 66(2): 117-126.
- Sivanesan A (1987): Graminicolous species of Bipolaris, Curvularia, Derechslera, Exserohilum and their teleomorphs. Mycological Papers 158:1-261, CAB International Mycological Institute, Ferry Lane, Kew, Surrey, UK.
- Sterflinger K (2010): Fungi: their role in deterioration of cultural heritage. Fungal Biology Reviews 24:47-55.
- Steiger M, Charola AE and Sterflinger K (2011): Weathering and deterioration. In: Architecture. Siegemud S and Snethlage R (eds.), Springer, Heidelberg, Germany, pp. 291-304.
- Urzi C and Realini M (1998): Colour changes of notos calcareous sandstone as related to its colonization by microorganisms. International Biodeterioration and Biodegradation 42: 45-54.
- Urzi C, De-Leo F, Paola S and Criseo G (2001): Air-borne fungal spores colonizing marbles exposed in the terrace of Messina Museum, Sicily. Aerobiologia 17(1): 11-17.
- Wanfu W, Xu M, Yantian M, Lin M, Fast W, Xiaojun M, Lizhe A and Huyuan F (2010): Seasonal dynamics of airborne fungi in different caves of the Mogao Grottoes Dunhuang, China. International Biodeterioration and Biodegradation 64(6): 461-466.
- Yang CS and Johanning E (1997): Airborne fungi and mycotoxins. In: Hurst CJ, Knudsen GR, McInerney MJ, Stetzenbach LD and Walter MV (Eds.): Manual of Environmental Microbioliogy, ASM Press, Washington, D.C., pp.651-660.
- Zielinska-Jankiewicz K, Kozajda A, Piotrowska M and Szadkowska-Stanczyk I (2008): Microbiological contamination with moulds in work environment in libraries and archive storage facilities. Annals of Agricultural and Environmental Medicine 15: 71-78.