Contribution to the mycobiota of Egypt

Neurospora tetrasperma Shear & Dodge; a new record to Egypt

A. H. Moubasher¹, A. M. Moharram and Zeinab Soliman

Assiut University Mycological Centre, Assiut University, Assiut, Egypt Received 28/1/2010, Accepted 20/2/2010

¹Corresponding author: e-mail: <u>moubasher29@yahoo.com</u>

Abstract: One ascosporic isolate of *Neurospora* was isolated during surveys of filamentous fungi of *Citrus* and grapevine plantations in Assiut area, Egypt. The isolate was examined for its macroscopic and microscopic features and identified as *Neurospora tetrasperma*. This species is being recorded here for the first time from Egypt.

Key words: Neurospora tetrasperma, Citrus plantation, Mycobiota, Egypt

Introduction

The generic name Neurospora was introduced by Shear and Dodge (1927) for four species characterized by dark ascospores, with a grooved surface with longitudinal ribs. Gelasinospora was proposed by Dowding (1933) to accommodate two species with ascospores similar to those of Neurospora, but with a pitted wall. Thereafter, many species of Neurospora and Gelasinospora, so far 12 species of Neurospora (Tai 1935, Gochenaur & Backus 1962, Nelson et al. 1964, Frederick et al. 1969, Mahoney et al. 1969, von Arx 1981, Perkins & Raju 1986, Krug & Khan 1991, Dettman et al. 2001) and 37 of Gelasinospora (refer to Garcia et al. 2004) have been described. Different members of these genera use one of three different mating strategies: heterothallism, homothallism, or pseudohomothallism. Recent studies on both Neurospora and Gelasinospora based on DNA sequences of four nuclear genes (Dettman et al. 2001), and ultrastructural and 28S rDNA sequence data of numerous strains of most of the species of both genera (Garcia et al. 2004) revealed that the ornamentation pattern of the epispore is not a very informative character for establishing phylogenetic relationships at the infrageneric level in these two genera and confirm the hypothesis that there are not enough criteria to distinguish them from each other. The names Neurospora and Gelasinospora are synonymized and so far 49 species of Neurospora have been now recognized in the genus (Garcia et al. 2004) and the generic diagnosis was expanded to include those with ascospores broadly fusiform, ellipsoidal, or nearly spherical, 1-celled, hyaline to yellowish brown or olive-brown, becoming dark and opaque at maturity, ascosporic wall with longitudinal ribs or pitted, occasionally nearly smooth. Anamorphs are known in only a relatively small number of species, belonging to Chrysonilia Arx 1981 (Garcia et al. 2004). However, investigations on multiple-gene sequences and morphology

concluded that although *Gelasinospora* and *Neurospora* are closely related, there is insufficient evidence to place currently accepted *Gelasinospora* and *Neurospora* species into the same genus (Cai *et al.* 2006).

In Egypt, of this genus only the anamorphic stage of *Neurospora crassa* (*Chrysonilia crassa*) is being reported. It has been reported from animal and bird pens materials (Moharram *et al.* 1987), soybean meal (Moharram *et al.* 1989), combine harvestor sorghum dust (Abdel-Hafez *et al.* 1990), caraway and cumin seeds (Abdel-Mallek *et al.* 1990), the air of Bahariya Oasis, Western Desert (Ismail *et al.* 2002). Reference strains of this species are deposited in the Culture Collection of Assiut University Mycological Centre (refer to the Catalogue of Culture Collection of AUMC 2010).

Neurospora tetrasperma C. L. Shear & B. O. Dodge 1927

Neurospora erythraea C. L. Shear & B. O. Dodge 1927

Neurospora toroi F. L. Tai 1935

Anamorph: *Chrysonilia tetrasperma* (C. L. Shear & B. O. Dodge) Arx 1981

Monilia tetrasperma Shear & Dodge 1927

Colony characteristics

Colonies on PSA and OA filling the plates (9 cm) within 3 days at 25 °C; ascomata produced abundantly after 7 days, dark brown to black, with scanty aerial hyphae and dark reverse particularly below the ascomata. Colonies on CYA filling the plate within 3 days, after 7 days, dark but intermixed with orange colour (the conidial stage) especially at the periphery of the plate, reverse is dark below the ascomata (but lighter than that on PSA and OA). On MEA after 7 days, colonies and reverse orange, with very little numbers of ascomata compared to those on PSA, OA and CYA (Fig. 1).

Teleomorph

Homothallic, ascomata ostiolate, 200-370 μ m in diam, beak length 58-110 μ m, beak width 80-88 μ m. Asci 4-spored, 107-131x15-17 μ m. Ascospores uniseriate or somewhat overlapping, olivaceous at first, becoming dark brown to black with maturity, ellipsoidal or elongate, 33.6-36x14-16 μ m, wall with 16–22 longitudinal, sometimes branched ribs and ornate epispore, with one germ pore at each end (Fig. 2).

Anamorph

Produced abundantly on MEA and CYA, conidia subglobose to obovoid, smooth, 8-12(-16) x4-9 μ m, orange in mass.

This species was first described by Shear and Dodge (1927) from an isolate contributed by Dr Gerald Stahel from unknown substratum. It is quite different from the other species producing 4-spored asci (*N. tetraspora* Garcia *et al.* 2004 = *Gelasinospora tetrasperma* Dowding 1933) by the ascospores dimensions $(21-33x12-20 \ \mu\text{m})$, and the inwardly projecting circular or slightly irregular pits of the ascospore wall (Garcia *et al.* 2004).

This is the first record of this species in Egypt. It is known by only one isolate and reference strain has been deposited in the Culture Collection of Assiut University Mycological Centre (AUMC 5089 recovered from the air of *Citrus* plantation in April 2008).

This species is of worldwide distribution. It has been frequently isolated from burnt vegetation and soil (Shear & Dodge 1927, Tai 1935, Perkins *et al.* 1976, Perkins & Turner 1988, Turner *et al.* 2001, Garcia *et al.* 2004 and Jacobson *et al.* 2004), and alkaline soil (Eliades *et al.* 2006).

Key to the two species of *Neurospora* so far recorded in Egypt (both with ascospores with longitudinal striations):

- Homothallic, asci 4-spored, ascospores 24– 40 x 14–22 μm, conidia 8-12(-16)x4-9 μm.....N. tetrasperma

References

- Abdel-Hafez SII, Moubasher AH, Shoreit AAM and Ismail MA (1990): Fungal flora associated with combine harvester wheat and sorghum dusts from Egypt. J Basic Microbiol 30(7): 467-479.
- Abdel-Mallek AY, Abdel-Hafez AII And Moharram AM (1990): Contribution to the mycoflora of caraway, coriander and cumin seeds in Egypt.

Bulletin of the Faculty of Science, Assiut University 19 (1-D): 1-15.

- Arx JA von (1981): The Genera of Fungi Sporulating in Pure Culture. J. Cramer, Vaduz, 3rd edition.
- Cai L, Jeewon R and Hyde KD (2006): Phylogenetic investigations of Sordariaceae based on multiple gene sequences and morphology. Mycological Research 110: 137 150.
- Catalogue of Culture Collection of Assiut University Mycological Centre (AUMC), Assiut, Egypt, 2nd edition, March 2010
- Dettman JR, Harbinski FM and Taylor JW (2001): Ascospore morphology is a poor predictor of the phylogenetic relationships of *Neurospora* and *Gelasinospora*. Fungal Genetics and Biology 34: 49–61.
- Dowding ES (1933): *Gelasinospora*, a new genus of pyrenomycetes with pitted spores. Canadian Journal Research 9: 294–305.
- Eliades LA, Cabello MN and Voget CE (2006): Contribution to the study of Alkalophilic and alkali-tolerant Ascomycota from Argentina. Darwiniana 44(1): 64-73.
- Frederick LF, Uecher FA and Benjamin CP (1969): A new species of *Neurospora* from soil of West Pakistan. Mycologia 61: 1077–1084.
- Garcia D, Stchigel AM, Cano J, Guarro J and Hawksworth DL (2004): A synopsis and recircumscription of *Neurospora* (syn. Gelasinospora) based on ultrastructural and 28S rDNA sequence data. Mycol. Res. 108 (10): 1119–1142.
- Gochenaur SE and Backus MP (1962): A new species of Neurospora from Wisconsin lowland soil. Mycologia 54:555–562.
- Ismail MA, Abdel-Hafez SII and Moharram AM (2002): Aeromycobiota of western desert of Egypt. African Journal of Science and Technology (AJST) 3 (1): 1-9.
- Jacobson DJ, Powell AJ, Dettman JR, Saenz GS, Barton MM, Hiltz MD, Dvorachek, JrWH, Louise Glass N, Taylor JW and Natvig DO (2004): *Neurospora* in temperate forests of western North America. Mycologia 96: 66–74.
- Krug JC and Khan RS (1991): A new homothallic species of *Neurospora* from Hungary. Mycologia 83: 829–832.
- Mahoney DP, Huang LH and Backus MP (1969): New homothallic Neurosporas from tropical soils. Mycologia 61: 264–274.
- Moharram AM, Bagy MMK and Abdel-Mallek AY (1987): Saprophytic fungi isolated from animal and bird pens in Egypt. Journal of Basic Microbiology 27 (7): 361-367.
- Moharram AM, Abdel-Gawad KM, Megalla SE and Mahmoud A-LE (1989): Fungal flora of poultry feedstuff ingredients. Journal of Basic Microbiology 29 (8): 491-499.

- Nelson AC, Novak RO and Backus MP (1964): A new species of *Neurospora* from soil. Mycologia 56: 384–392.
- Perkins DD, and Raju NB (1986): *Neurospora* discreta, a new heterothallic species defined by its crossing behaviour. Experimental Mycology 10:323–338.
- Perkins DD and Turner BC (1988): *Neurospora* from natural populations: toward the population biology of a haploid eukaryote. Experimental Mycology 12: 91–131.
- Perkins DD, Turner BC and Barry EG (1976): Strains of *Neurospora* collected from nature. Evolution 30: 281–313.
- Shear CL and Dodge BO (1927): Life histories and heterothallism of the red bread-mold fungi of the *Monilia sitophila* group. Journal of Agricultural Research 34: 1019–1042.
- Tai FL (1935): Two new species of *Neurospora*. Mycologia 27: 328–330.
- Turner BC, Perkins DD and Fairfield A (2001): *Neurospora* from natural populations: a global study. Fungal Genetics and Biology 32: 67–92.

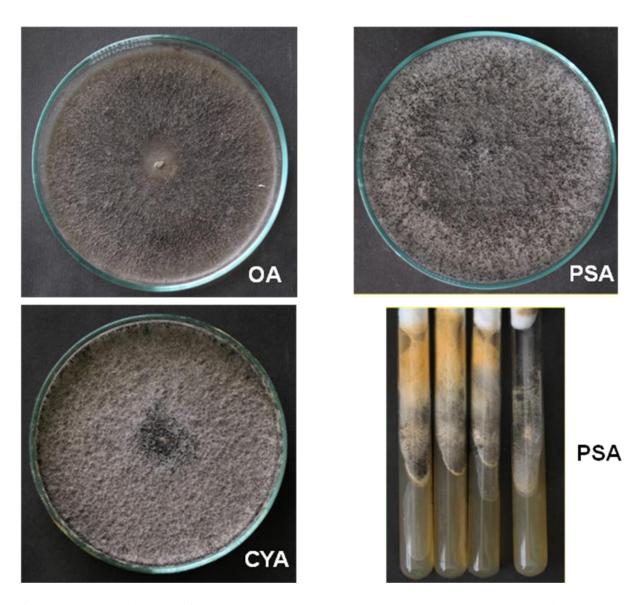


Figure 1: 15-day old cultures of *Neurospora tetrasperma* AUMC 5089 on Oat agar (OA), Czapek yeast agar (CYA), potato sucrose agar (PSA) plates and PSA slants.

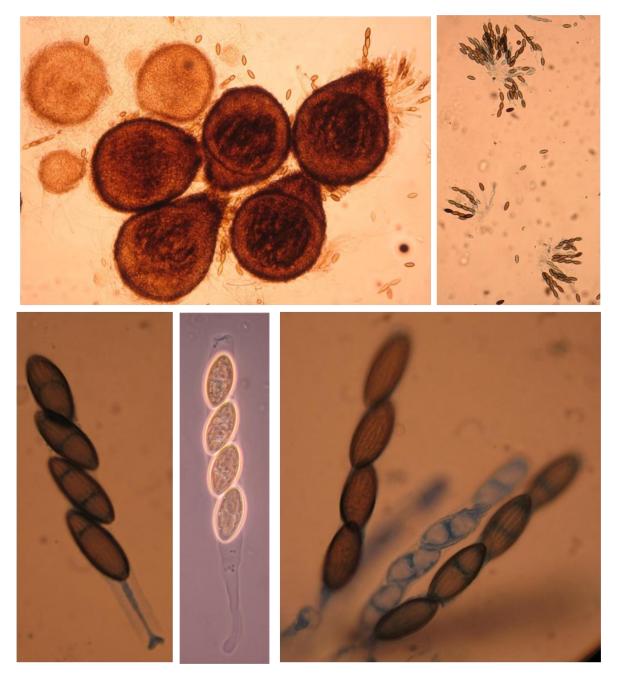


Figure 2: Ascomata, 4-spored asci and longitudinally striated ascospores of *Neurospora tetrasperma* AUMC 5089.